

GENETIC ARCHITECTURE FOR EARLINESS IN COTTON (*Gossypium hirsutum* L.)

*VALU, M. G.; PATEL, J. A.; POLARA, A. M.; MADARIYA, R. B. AND VARIYA, M. V.

COTTON RESEARCH STATION
JUNAGADH AGRICULTURAL UNIVERSITY
JUNAGADH-362 001 (GUJARAT), INDIA

**Email: mgvalu@jau.in*

ABSTRACT

*The experimental materials consisted of twelve generations, namely P_1 , P_2 , F_1 , F_2 , B_1 , B_2 , B_{11} , B_{12} , B_{21} , B_{22} , B_{1s} and B_{2s} of two crosses of cotton viz., Deviraj x GBHV-170 (cross-1) and G.Cot-10 x MR-786 (cross-2) with a view to generate genetic information on gene effects for earliness in cotton (*Gossypium hirsutum* L.). Special scaling tests such as X and Y were significant either in cross-1 or cross-2 for all the four traits besides significance of other tests showing presence of epistasis. The $X^2_{(2)}$ value at six degrees of freedom were significant in all the traits in both the crosses supported the presence of higher order epistasis. The $X^2_{(3)}$ value at two degrees of freedom was non-significant in cross-1 for number of monopodia per plant and in cross-2 for seed cotton yield per plant and days to 50 per cent boll bursting proving the ten parameter model as the best fit model. The $X^2_{(3)}$ value at two degrees of freedom was significant for seed cotton yield per plant, days to flowering traits in cross-1 and days to flowering and number of monopodia per plant in cross-2 indicating the presence of higher order epistasis and/or linkage.*

KEY WORDS: Cotton, Digenic, Gene Effects, Trigenic

INTRODUCTION

India is the only country where all the four cultivated species of cotton are grown on commercial scale and covers cultivated area about 105 lakh ha. It occupies second position in production with 351 lakh bales among all cotton producing countries, next to China. Average productivity of India is 568 kg/ha, which is much lower as compared to the world average productivity of 766 kg/ha. Gujarat is the second largest cotton growing state with acreage of 24 lakh ha and the largest cotton producing state of India with production of 95 lakh bales. The average productivity of cotton in the state is 673 kg/ha, which is higher than national productivity

(Anonymous, 2016). Earliness is desirable in cotton to allow escape from later stage infestation of insects pests and loss of yield under rainfed situation. The information on gene action for earliness is very essential for deciding the effective selection method in segregating generations. The additive and dominance gene effects may have great value on the improvement of seed cotton yield with earliness. The information on epistatic gene effect is also important for the yield improvement in cotton. Hence, the present investigation was under taken to study the gene action of earliness in cotton.

MATERIALS AND METHODS

The experimental materials consisted of twelve generations, namely P_1 , P_2 , F_1 , F_2 ,

B₁, B₂, B₁₁, B₁₂, B₂₁, B₂₂, B_{1s} and B_{2s} of two crosses of cotton viz., Deviraj x GBHV-170 (cross-1) and G.Cot-10 x MR-786 (cross-2). Experiment was laid-out in Compact Family Block Design with three replications during *Kharif* 2013 at Cotton Research Station, Junagadh Agricultural University, Junagadh. Each replication was divided into two compact blocks each consists of single cross and blocks were consisted of twelve plots comprised of twelve basic generations of each cross. The crosses were assigned to each block and twelve generations of a cross were randomly allotted to individual plot within the block. The plots of various generations contained different number of rows i.e., parents and F₁ in single row; B₁ and B₂ in two rows and F₂, B_{1s}, B₁₁, B₁₂, B_{2s}, B₂₁ and B₂₂ in three rows. Each row was of 6.3 m in length with 120 cm and 45 cm inter and intra row spacing, respectively. All the recommended agronomical practices and necessary plant protection measures were followed timely to raise good crop of cotton. The observations were recorded on seed cotton yield per plant, days to flowering, days to 50 per cent boll bursting and number of monopodia per plant on five randomly selected plants in each replication for P₁, P₂ and F₁; ten plants for B₁ and B₂ and twenty plants for F₂, B₁₁, B₁₂, B₂₁, B₂₂, B_{1s} and B_{2s}. To decide the adequacy of three, six and ten parameter model, simple scaling tests given by Hayman and Mather (1955), Hill (1966) and Van Der Veen (1959) were employed. Joint scaling test of Cavalli (1952) was applied to test adequacy of three, six and ten-parameter models. Whenever, this simple additive-dominance model failed to explain the variation in generation means, six and ten-parameter models using weighted least square method were used to estimate main, digenic and trigenic effects.

RESULTS AND DISCUSSION

The data were initially subjected to simple scaling tests A, B, C and D. Significant estimates of any one or more of these tests indicate the presence of digenic interactions. Further, simple scaling tests B₁₁, B₁₂, B₂₁, B₂₂, B_{1s} and B_{2s} given by Hill (1966) and X and Y given by Van Der Veen (1959) were also computed. The significant estimate of the test(s) given by Hill (1966) showed the contribution of particular generation to higher order epistasis which indirectly indicating the presence of epistasis. If any of the Van Der Veen's tests deviate significantly from zero indicates the presence of trigenic or higher order epistasis. The results of simple scaling tests were further confirmed by joint scaling test (Cavalli, 1952), which effectively combines the whole set of simple scaling tests. Thus, it offers a more general, convenient, adoptable and informative approach for estimating gene effects and also for testing adequacy of additive-dominance model. The $\chi^2_{(1)}$ test at nine degrees of freedom; $\chi^2_{(2)}$ at six degrees of freedom and $\chi^2_{(3)}$ at two degrees of freedom were applied to test the fitness of three-parameter model, six-parameter model and ten-parameter model, respectively. The ten-parameter model was used to estimate higher order epistasis (Hill, 1966). To draw inference on adequacy of ten-parameter model, chi-square test $\chi^2_{(3)}$ at two degrees of freedom was applied. The degree of freedom for χ^2 was computed by subtracting number of parameters considered under the respective model from the number of generations. The results are presented in Table 1 and 2.

Out of all the scaling tests, only A, B, C, D and B₂₁ in cross-1 and A, B, C, B₁₂, B₂₁ and special scaling test Y in cross-2 were significant showing presence of epistasis for seed cotton yield per plant, while all the scaling tests except B₂₂ in cross-1 and all the scaling tests except B, C,

B₁₁ and B₂₅ cross-2 were significant showing presence of digenic and trigenic gene action for days to flowering. For days to 50 per cent boll bursting, the scaling tests A, C, D, B₁₁, B₁₂, B₂₁, B₂₂, B₁₅ and X in cross-1 and scaling tests A, B, C, B₁₂, B₂₁, B₂₅, X and Y in cross-2 were significant showing presence of epistasis. On the other hand, the scaling tests A, B, C, B₁₁, B₁₂, B₂₁, B₁₅, X and Y in cross-1 and A, B, C, B₁₁, B₁₂, B₂₁, B₂₂, B₁₅, B₂₅ and X in cross-2 were significant showing presence of digenic and trigenic gene interaction for number of monopodia per plant. All the three parameters i.e. 'm', additive [d] and dominance [h] of three parameter model were significant in cross-1 and cross-2 for all the characters under study, except additive [d] in cross-2 for seed cotton yield per plant and dominance [h] in cross-1 for seed cotton yield per plant. The $X^2_{(1)}$ values with nine degrees of freedom of joint scaling test was significant in all the characters indicating the failure of additive-dominance model which indirectly pointed out the presence of epistasis. Cockerham (1959) postulated that the epistatic gene action is common in the inheritance of quantitative traits and there is no sound biological reason why this type of gene action should be less common for these traits.

When the simple additive-dominance model failed to explain the variation among generation means, a six parameter model involving three digenic interactions ([i], [j] and [l]) based on weighted least square technique proposed by Hill (1966) was tested which had provision of testing the adequacy of model with six degrees of freedom besides being utilizing means of all the twelve generations. Hence, the present study was planned to execute with means of twelve generations and model of Hill (1966) was tested in which six degrees of freedom left for testing the adequacy of six parameter model of Hill (1966). According to the six

parameter model of Hill, the parameters 'm', [d] and digenic [i] in cross-1 and all the parameters except digenic [j] in cross-2 were significant for seed cotton yield per plant, while all the parameters in cross-1 and except digenic [j] in cross-2 were significant for days to flowering. Likewise, for days to 50 per cent boll bursting, the estimate of 'm', [h], [i], [j] and [l] in cross-1 and 'm', [d], [h], [j] and [l] in cross-2 were significant, while all the estimate of gene effects except [d] and [i] in cross-1 and [i] in cross-2 were significant for number of monopodia per plant. The $X^2_{(2)}$ value at six degrees of freedom were significant in all four traits in two crosses indicating the presence of higher order epistasis.

In ten parameter model, dominance x dominance [l] and dominance x dominance x dominance [z] were significant in both the crosses for seed cotton yield per plant and additionally dominance [h], additive x additive [i] and additive x additive x dominance [x] in cross-1 and 'm' in cross-2. For days to flowering, 'm', additive x dominance [j], dominance x dominance [l], additive x dominance x dominance [y] and dominance x dominance x dominance [z] were found significant in both the crosses, and additionally dominance [h], additive x additive [i] and additive x additive x dominance [x] in cross-1 and additive [d] and additive x additive x additive [w] in cross-2. The 'm', additive x dominance [j], additive x dominance x dominance [y] and dominance x dominance x dominance [z] were found significant in both the crosses for days to 50 per cent boll bursting, additionally dominance [h], additive x additive [i], dominance x dominance [l] and additive x additive x dominance [x] in cross-1 and additive x additive x additive [w] in cross-2. For number of monopodia per plant, the gene effects additive x dominance x dominance [y] and dominance x dominance x dominance [z] were significant in cross-1,

while 'm', dominance [h], additive x additive [i], dominance x dominance [l], additive x additive x additive [w], additive x additive x dominance [x] and dominance x dominance x dominance [z] were significant in cross-2. The $X^2_{(3)}$ value at two degrees of freedom was non-significant for seed cotton yield per plant and days to 50 per cent boll bursting in cross-2 and number of monopodia per plant in cross-1 depicting that the ten parameter model as the best fit model. The $X^2_{(3)}$ value at two degrees of freedom was significant in all the traits under study except number of monopodia per plant in cross-1 and for seed cotton yield per plant and days to 50 per cent boll bursting in cross-2, indicating the presence of higher order epistasis and/or linkage.

These findings were further confirmed from the investigations done by several researchers, who worked on different kind of gene effects mostly up to digenic interactions and there is no report on trigenic interactions in cotton so far. However, few reports are available in different crops viz., Bhapkar and D'Cruz (1967) and Singh (2012) in castor and Sharma *et al.* (2002) in wheat. The opposite signs of either two or all the three gene effects viz., dominance [h], dominance x dominance [l] and dominance x dominance x dominance [z] suggested the presence of duplicate type of epistasis. In present study, duplicate epistasis was observed in both the crosses for all the four traits under investigation. Duplicate type of epistasis also reported by Thombre *et al.* (1987) for seed cotton yield per plant; by Mehetre (2003) for days to boll bursting, seed cotton yield per plant and number of monopodia per plant; by Esmail (2007) for days to first flowering and seed cotton yield per plant; by Haleem *et al.* (2010) for days to flowering seed cotton yield and by Kannan *et al.* (2013) for single plant yield.

CONCLUSION

From the foregoing discussions, it could be concluded that earliness recorded in two crosses were governed by additive, dominance and digenic and/or trigenic epistasis gene effects along with duplicate type of gene action. When additive as well as non-additive gene effects are involved, a breeding scheme efficient in exploiting both types of gene effects should be employed. Bi-parental mating could be followed which would facilitate exploitation of both additive and non-additive gene effects simultaneously for genetic improvement of seed cotton yield with earliness in cotton.

REFERENCES

- Anonymous. (2016). All India Coordinated Cotton Improvement Project. Annual Report, CICR, Regional station, Coimbatore.
- Bhapkar, D. G. and D'Cruz, R. (1967). Inheritance of oil content in *Ricinus communis*. *Indian J. Genet.*, **27**: 152-153.
- Cavalli, L. L. (1952). An Analysis of Linkage in Quantitative Inheritance. In "*Quantitative Inheritance*". Ed. E.C.R. Reeve and C. H. Waddington, HMSO, London. pp. 135-144.
- Cockerham, C. C. (1959). Partition of hereditary variance for various genetic models. *Genetics*, **44**(6): 1141-1148.
- Esmail, R. M. (2007). Genetic analysis of yield and its contributing traits in two intra specific cotton crosses. *J. Appl. Sci. Res.*, **3**(12): 2075-2080.
- Haleem, S. H. M. A.; Metwali, E. M. R and Felail, A. M. M. (2010). Genetic analysis of yield and its component in some Egyptian cotton (*Gossypium barbadense* L.) varieties. *World J. Agric. Sci.*, **6**(5): 615-621.
- Hayman, B. I. and Mather, K. (1955). The description of genetic interactions in

- continuous variation. *Biometrics*, **11**(1): 69-82.
- Hill, J. (1966). Recurrent backcrossing in the study of quantitative inheritance. *Heredity*, **21**: 85-120.
- Kannan, S.; Ravikesavan, R. and Govindaraj, M. (2013). Genetic analysis for quantitative and quality characters in three single crosses of upland cotton. *Nat. Sci. Biol.*, **5**(4): 450-453.
- Mehetre, S. S.; Shinde, S. K. and Shinde, G. C. (2003). Genetic analysis for seed cotton yield and its components in cotton. *J. Maharashtra Agric. Univ.*, **28**(1): 46-50.
- Sharma, S. N.; Sain, R. S. and Sharma, R. K. (2002). Genetic control of quantitative traits in *durum* wheat under normal and late sowing environments. *SABRAO J. Breed. Genet.*, **34**(1): 35-43.
- Singh, A. P. (2012). Gene systems for seed yield and its component traits in castor (*Ricinus communis* L.). Ph. D. (Agri.), Thesis (unpublished) submitted to JAU, Junagadh.
- Thombre, M. V.; Pathade, S. V. and Patil, F. B. (1987). Genetic analysis for boll number, boll weight and seed cotton yield. *J. Maharashtra Agric. Univ.*, **12**(3): 306-308.
- Van Der Veen, J. H. (1959). Test of non-allelic interaction and linkage for quantitative character in generation derived from two diploid pure lines. *Genetica*, **30**(1): 201-232.

Table 1: Scaling tests and estimation of gene effects for seed cotton yield per plant and days to flowering in two crosses of cotton

Scaling Tests /Gene Effects	Seed Cotton Yield Per Plant				Days to Flowering			
	Deviraj x GBHV-170 (cross 1)		G.Cot-10 x MR-786 (cross 2)		Deviraj x GBHV-170 (cross 1)		G.Cot-10 x MR-786 (cross 2)	
A	21.13**	± 6.94	-37.73**	± 9.96	22.53**	± 0.94	1.67*	± 0.76
B	35.47**	± 9.22	-24.00**	± 6.34	3.67**	± 1.16	1.33	± 0.75
C	99.73**	± 12.03	-40.07**	± 13.65	8.07**	± 1.66	-3.00	± 1.67
D	21.57**	± 7.90	10.83	± 8.43	-9.07**	± 0.87	-3.00**	± 0.86
B₁₁	-1.00	± 17.07	-8.40	± 16.92	-48.13**	± 1.56	-1.47	± 1.57
B₁₂	12.27	± 17.77	65.33**	± 18.96	-33.73**	± 1.97	-7.20**	± 1.98
B₂₁	47.07**	± 13.44	84.93**	± 16.84	-35.20**	± 1.97	5.20**	± 1.74
B₂₂	14.67	± 21.45	0.53	± 11.18	1.33	± 2.08	8.20**	± 1.93
B_{1s}	8.53	± 35.69	-8.67	± 35.84	-83.73**	± 3.20	10.67**	± 3.16
B_{2s}	-3.87	± 36.79	-2.67	± 31.76	-23.60**	± 4.00	0.40	± 3.43
X	-12.62	± 8.51	-7.13	± 7.59	-12.00**	± 0.71	-5.52**	± 0.79
Y	11.42	± 8.67	39.53**	± 7.84	-5.53**	± 0.85	-2.18*	± 0.85
Three Parameter Model								
m	120.58**	± 1.09	98.89**	± 1.25	77.97**	± 0.18	73.87**	± 0.16
(d)	7.53**	± 1.11	1.96	± 1.26	3.72**	± 0.17	1.40**	± 0.16
(h)	22.31**	± 1.93	33.29**	± 2.30	2.15**	± 0.38	-1.80**	± 0.32
$\chi^2_{(1)} (9 \text{ df})$	112.35**		60.06**		1632.27**		85.38**	
Six Parameter Model								
m	142.14**	± 9.54	123.05**	± 8.93	71.72**	± 0.86	70.95**	± 0.85
(d)	8.58**	± 1.19	2.97*	± 1.46	1.67**	± 0.26	1.61**	± 0.20
(h)	1.16	± 24.88	-54.17*	± 22.96	33.59**	± 2.44	5.29*	± 2.24
(i)	-24.29*	± 9.56	-20.84*	± 8.91	3.60**	± 0.85	3.03**	± 0.86
(j)	-15.36	± 7.91	-12.80	± 7.75	14.44**	± 0.96	-1.48	± 0.80
(l)	-3.93	± 16.28	68.26**	± 15.11	-31.71**	± 1.87	-4.30**	± 1.58
$\chi^2_{(2)} (6 \text{ df})$	74.84**		31.53**		775.14**		69.59**	
Ten Parameter Model								
m	-15.61	± 26.61	91.11**	± 24.78	111.50**	± 2.26	76.32**	± 2.52
(d)	-3.19	± 22.99	3.11	± 20.05	-2.97	± 1.78	-8.20**	± 1.88
(h)	789.65**	± 128.89	155.71	± 123.72	-178.67**	± 11.57	-24.26	± 13.11
(i)	133.82**	± 26.63	9.12	± 24.79	-35.63**	± 2.27	-2.54	± 2.53
(j)	48.23	± 62.14	9.34	± 51.53	55.42**	± 4.87	33.13**	± 5.21
(l)	-1163.29**	± 194.52	-365.87*	± 178.16	288.46**	± 17.95	45.07*	± 20.21
(w)	11.32	± 22.98	-0.70	± 20.01	2.52	± 1.77	9.51**	± 1.88
(x)	-432.56**	± 66.32	-44.63	± 68.20	122.89**	± 6.47	12.53	± 7.33
(y)	-81.78	± 58.29	-34.94	± 49.17	-68.15**	± 4.68	-38.61**	± 5.26
(z)	529.35**	± 93.31	258.79**	± 90.35	-148.74**	± 8.90	-25.63**	± 9.78
$\chi^2_{(3)} (2 \text{ df})$	27.12**		1.10		66.64**		12.78**	
Type of Epistasis	Duplicate		Duplicate		Duplicate		Duplicate	

*, ** Significant at 5 and 1 per cent levels, respectively

Table 2: Scaling tests and estimation of gene effects for days to 50 per cent boll bursting and number of monopodia per plant in two crosses of cotton

Scaling Tests /Gene Effects	Days to 50 Per Cent Boll Bursting		Number of Monopodia Per Plant	
	Deviraj x GBHV-170 (cross 1)	G.Cot-10 x MR-786 (cross 2)	Deviraj x GBHV-170 (cross 1)	G.Cot-10 x MR-786 (cross 2)
A	17.40** ± 2.34	8.07** ± 1.69	-2.07** ± 0.47	2.53** ± 0.42
B	2.80 ± 1.95	9.47** ± 1.74	-1.40** ± 0.43	1.33** ± 0.42
C	8.60* ± 3.38	21.27** ± 3.41	-2.93** ± 0.83	3.00** ± 0.75
D	-5.80** ± 1.96	1.87 ± 1.85	0.27 ± 0.43	-0.43 ± 0.35
B₁₁	-45.40** ± 3.54	-4.27 ± 3.31	3.93** ± 0.83	-6.40** ± 0.78
B₁₂	-34.20** ± 3.39	-25.73** ± 2.63	4.07** ± 0.89	-4.27** ± 0.87
B₂₁	-32.73** ± 3.56	-11.53** ± 2.86	4.87** ± 0.96	-5.47** ± 0.85
B₂₂	-10.20* ± 3.97	-1.67 ± 3.54	-0.27 ± 0.89	-2.47** ± 0.71
B_{1s}	-53.87** ± 7.08	-11.60 ± 6.82	6.07** ± 1.40	-12.67** ± 1.49
B_{2s}	-12.40 ± 7.31	-28.80** ± 6.27	1.40 ± 1.50	-3.33* ± 1.39
X	-9.17** ± 1.49	-4.20** ± 1.28	0.85* ± 0.36	-0.68* ± 0.27
Y	-2.83 ± 1.65	-7.83** ± 1.41	1.32** ± 0.40	-0.22 ± 0.36
Three Parameter Model				
m	113.08** ± 0.36	112.39** ± 0.33	1.62** ± 0.08	1.50** ± 0.07
(d)	1.87** ± 0.35	1.47** ± 0.32	-0.24** ± 0.08	-0.22** ± 0.07
(h)	-0.95 ± 0.69	-4.31** ± 0.59	0.67** ± 0.17	0.65** ± 0.14
$\chi^2_{(1)} (9 \text{ df})$	293.77**	131.96**	55.45**	118.27**
Six Parameter Model				
m	100.57** ± 1.83	110.50** ± 1.65	2.58** ± 0.37	0.72* ± 0.35
(d)	0.86 ± 0.48	2.52** ± 0.43	-0.06 ± 0.12	-0.45** ± 0.09
(h)	46.12** ± 5.05	10.99* ± 4.39	-3.60** ± 1.07	4.96** ± 1.01
(i)	10.26** ± 1.85	-0.34 ± 1.68	-0.57 ± 0.37	0.38 ± 0.35
(j)	10.59** ± 1.87	-5.93** ± 1.54	-0.99* ± 0.44	1.37** ± 0.33
(l)	-39.53** ± 3.67	-16.42** ± 3.15	3.97** ± 0.83	-4.57** ± 0.81
$\chi^2_{(2)} (6 \text{ df})$	123.93**	48.10**	19.75**	48.51**
Ten Parameter Model				
m	126.97** ± 5.00	115.23** ± 4.79	0.62 ± 1.07	3.74** ± 0.93
(d)	-4.62 ± 3.92	-7.02 ± 3.59	0.18 ± 0.74	1.32 ± 0.73
(h)	-94.49** ± 25.55	-17.45 ± 24.76	7.87 ± 5.76	-11.04* ± 4.80
(i)	-15.80** ± 5.01	-4.17 ± 4.80	1.27 ± 1.07	-2.70** ± 0.94
(j)	48.61** ± 10.75	27.02** ± 9.69	-3.73 ± 2.23	-0.81 ± 1.99
(l)	171.73** ± 38.99	44.04 ± 37.20	-16.38 ± 9.06	19.05* ± 7.44
(w)	4.20 ± 3.90	9.10* ± 3.57	-0.10 ± 0.74	-1.91** ± 0.73
(x)	80.82** ± 14.36	-2.88 ± 14.35	-4.46 ± 3.29	9.87** ± 2.67
(y)	-56.39** ± 10.06	-32.32** ± 8.68	4.75* ± 2.32	-1.14 ± 1.83
(z)	-97.66** ± 18.80	-38.20* ± 17.49	11.13* ± 4.47	-10.70** ± 3.74
$\chi^2_{(3)} (2 \text{ df})$	44.53**	2.35	2.95	16.26**
Type of Epistasis	Duplicate	Duplicate	Duplicate	Duplicate

*, ** Significant at 5 and 1 per cent levels, respectively

[MS received : June 16 , 2018]

[MS accepted : June 22, 2018]